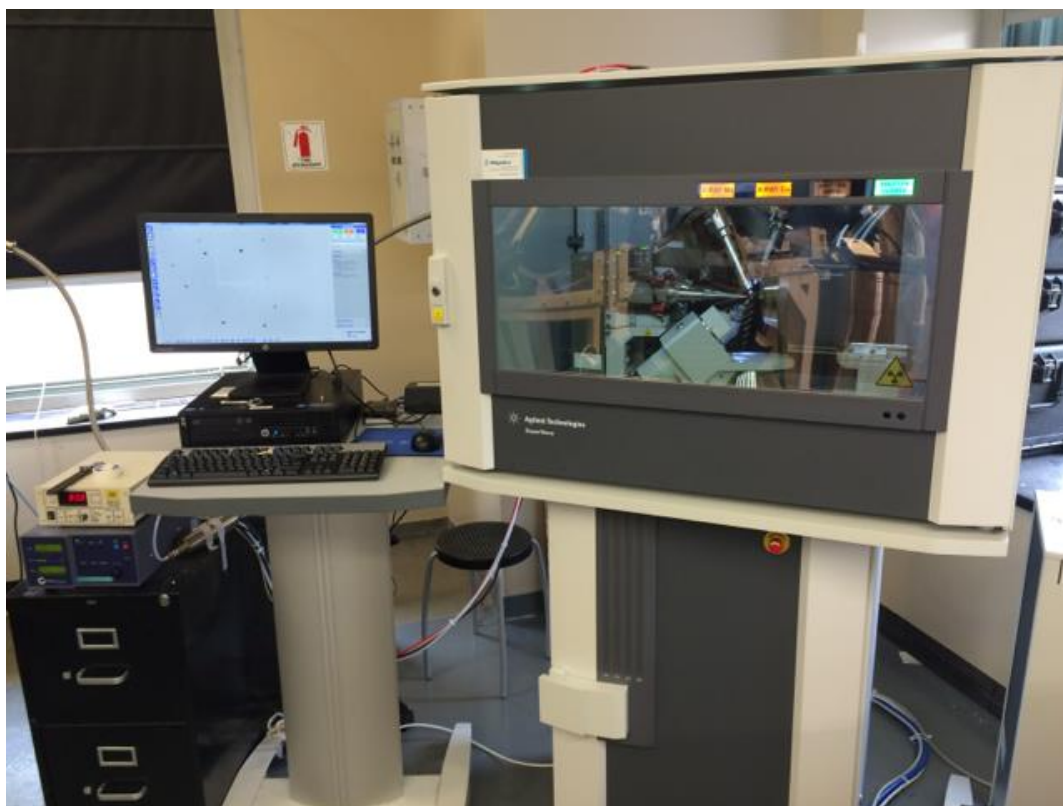




Agilent SuperNova SCXRD Standard Operating Procedure



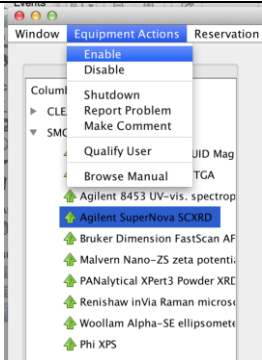
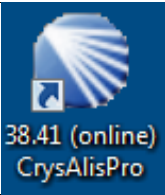
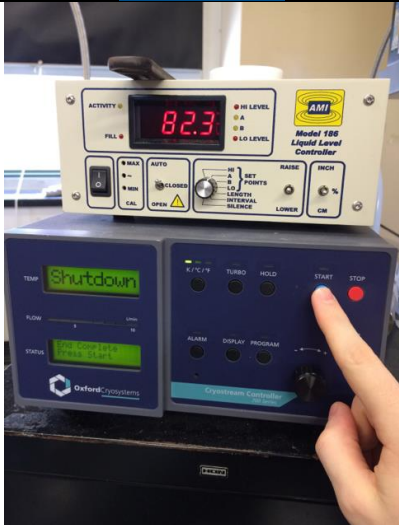
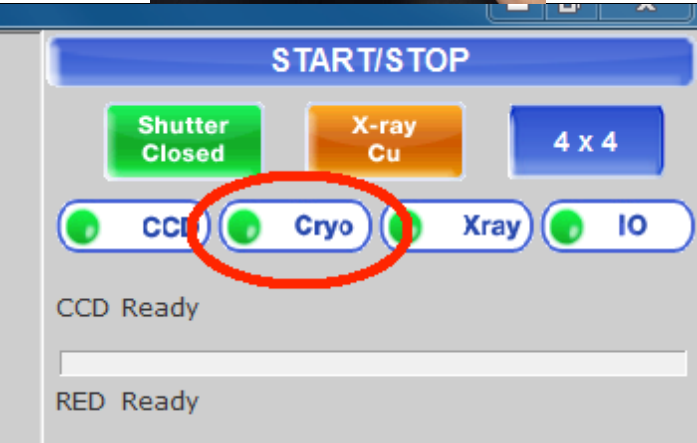
These instructions are intended for reference only, and will *not* replace the thorough training required for proper system operation. Contact staff/superuser with questions or to report a system problem.

Written by Daniel Paley.

Updated by Manju Rajeswaran (Oct, 2021)





<p>1.</p>	<p>Enable the tool in BADGER</p>	
<p>2.</p>	<p>If no window reading “CrysAlisPro (online)” is open, then start one from the desktop.</p>	
<p>3.</p>	<p>If the Cryostream is not running (display reads “Shutdown”), then press START and wait 10 seconds to initialize.</p>	
<p>4.</p>	<p>Click “Cryo.” Click “Set,” “Cool,” enter 100 K, click OK. From room temperature, the Cryostream takes about 20 min to cool down.</p>	





<p>5.</p>	<p>Click “Xray.” Click “Set kV,mA,X-ray”. Choose Auto-ramp and click OK.</p>	
<p>6.</p>	<p>Prepare a microscope slide with 1 drop of oil. Do not spill. Turn on the light.</p>	
<p>7.</p>	<p>Transfer a few crystals to the oil.</p>	





<p>8.</p>	<p>Use a knife to separate a small, single-crystalline fragment.</p> <p>The best crystals are 0.05 to 0.2 mm in size and regularly shaped. The full scale bar is 10 mm (1x zoom); 2.5 mm (4x zoom).</p> <p>Push the crystal out of the oil droplet so you can pick it up easily.</p>	
<p>9.</p>	<p>Pick a goniometer tip. Use your oil droplet to clean it off, then remove all excess oil by touching the glass.</p> <p>Mount the crystal neatly and without any excess oil. The crystal should be easily visible so you can center it on the diffractometer.</p>	
<p>10.</p>	<p>In CrysAlis, click “Start/Stop” and select “Start new.” Click “Mount.”</p>	





<p>11. Open the cabinet with the large key. Using your right hand, place the magnetic crystal mount on the goniometer. Do not touch anything besides the goniometer. The X-ray sources, camera and Cryostream head are carefully aligned; the detector window is made of toxic beryllium.</p>	
<p>12. In the mounting window, set phi=180.</p>	
<p>13. Using the small key and your right hand, adjust the crystal up/down with the upper screw and left/right with the lower screw.</p>	





<p>14. Rotate to $\phi=90$ and adjust the crystal left/right again.</p>	
<p>15. Rotate the ϕ axis to 2 positions separated by 90 degrees and ensure the crystal is well centered. The x-ray beam is half the diameter of the circle on the video display.</p>	
<p>16. Exit the mounting window and close the cabinet. Click the small arrow next to "Screening." Set $\theta=-35$, exposure time about 2-5 seconds. Click "Ok & Screen."</p>	





<p>17. Evaluate the diffraction. The spots should be round and reasonably intense with sharp edges. If the diffraction is not good, you can screen more crystals.</p>																					
<p>18. If the diffractometer is still cooling down, wait for it to reach 100 K. Use the “Mount” window to ensure your crystal is still centered.</p>																					
<p>19. Use the pre-experiment slider to set an appropriate frame time. Click “Start Pre-Exp.” This will collect 30 frames in 6 different positions. The wide angle frames are exposed for 5x the time you select.</p>																					
<p>20. When the pre-experiment finishes, a strategy window will launch. Set the wide-angle exposure time for predicted individual I/sigma around 6 to 10 and the small-angle time for 1/5 of the wide angle. Set “Scan width” to 1.</p>	<table border="1"> <thead> <tr> <th>Theta Range</th> <th>exp time (uncorrelated)</th> <th>individual I/sigma</th> <th>merged I/sigma</th> <th>theta binning</th> </tr> </thead> <tbody> <tr> <td>[-8.50; 35.53]</td> <td>2</td> <td>9.57</td> <td>15.13</td> <td>4x4H</td> </tr> <tr> <td>[-8.50; 108.00]</td> <td>10</td> <td>7.67</td> <td>16.94</td> <td>4x4H</td> </tr> <tr> <td>Total</td> <td></td> <td>7.97</td> <td>16.63</td> <td></td> </tr> </tbody> </table> <p>Predicted resolution beyond 0.82 Scan width: 1.00 Use theta-dependent binning/SSC <input checked="" type="checkbox"/></p>	Theta Range	exp time (uncorrelated)	individual I/sigma	merged I/sigma	theta binning	[-8.50; 35.53]	2	9.57	15.13	4x4H	[-8.50; 108.00]	10	7.67	16.94	4x4H	Total		7.97	16.63	
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<p>21. Under Strategy Parameters, choose “Other,” “hemisphere.” Check that “Resolution” is set to 0.8. Use “Complete Data” mode with 100% completeness. Click “Calculate Strategy.”</p>	
<p>22. To stop cryo after data collection, In strategy parameters - select Autochem/ Movie/Cryo/Red</p>	
<p>23. In the experiment options window select “Auto cryo/hot device shutdown on experiment completion option”. Click OK.</p>	





<p>24.</p>	<p>Click “Start named experiment.” Use “Browse” to find your data folder. Enter a name (preferably a notebook page #.) Enter the elements expected in your crystal. Click Start.</p>	
<p>25.</p>	<p>Ensure that the microscope light is off and the area is clean. If you leave samples, they will be thrown away.</p>	
<p>26.</p>	<p>Adjust your tool reservation in badger with your predicted finishing time.</p>	
<p>27.</p>	<p>When your experiment is finished, you can use Olex2 to solve your structure.</p>	





<p>28. When you are done, before you disable, please check to make sure that X-ray power is down to kV: 12.00 mA: 0.05 or it is ramping down. If not, please ramp down X-ray power manually.</p>	
<p>29. Select "X-ray"</p>	
<p>30. Click on "Set kV,mA,X-ray".</p>	





<p>31. Select “Standby” and “OK”</p>	
<p>32. BADGER LOGOUT: Disable the SCXRD in Badger.</p>	

