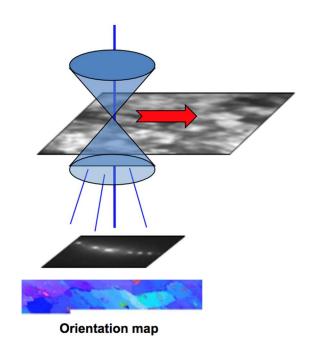
ASTAR (Orientation and Phase Mapping System on TEM)

Standard Operating Procedure



These instructions are intended for reference only, and will *not* replace the thorough training required for proper system operation. Contact a clean room staff member with questions or to report a system problem.



1.	Enable the TEM in BADGER	The ASTAR PC with black case on the ground (right side) should be on. Otherwise turn the PC on.
2.	Turn on the NanoMEGAS controller (behind the big monitor on the right)	(the switch is on the back)
3.	After loading the sample, find the region of interest and adjust the eucentric height (the <i>Z</i>)	
4.	Load the FEG Registry 200kv_TEM-NanoMegas	Set Update Delete Lbl Date 200kv_TEM 6/5/2018 80kv_TEM 11/9/2016 80kv_STEM 11/9/2016 200kv_TEM-NanoMeg 5/25/2018 200kv_TEM-NanoMeg 5/25/2018



5.	Tune the beam:	
J.	Gun shift,	At this beam setting, the beam is significantly
	Beam tilt pp x	less bright and less invasive.
	Beam tilt pp y	
	Beam shift	
6.	Fine tune the eucentric height	Adjusting the Z height is crucial in the final
	(adjusting the Z) by converging	image quality
	the beam (intensity) to a point.	
	The goal is that the beam	
	produces the minimum amount	
	of diffracting spots.	
7.	Click on the pause button on the	
	top menu of FluCam screen	
	Co to the Negal Acces to by front	
8.	Go to the NanoMegas tab (next	
	to the tabs STEM, Tune, etc.)	
	And click on both grey buttons (NanoMegas and FluCam)	
0	On the computer on the right	
9.	open the application Topspin	$\overline{\mathbf{x}}$
	Data Collection	
		DigiStar Topspin Data Topspin Data Control Collection Viewer
10.	In the first window, enter you	
	sample name.	
	The window should look like	VISIN Das Dilactos (pulsatos planatos parates tentos tento
11.	this. Click on the search button.	T PLD STDM Prot Point Diff From Series Area Free Device From Series Diff. Series
	By using the track ball , bring the	Lacol a TLU Survey Lacol Logo Logo Logo Logo Logo Logo Logo Lo
	beam to the center.	Insent Time () (3.1
		Forme Films The max Table A and A a
		France Date (1) 2 (1) (1) (1) (1) (1) (1) (1) (1) (1) (1)
		Sea Rago A
		So (mark the s) Anexa Tane () (200)
12.	Use the intensity knob , make	Tel dependence 4000 in the face and the face
12.	the beam to a spot.	

CNI Shared Facilities

13.	From the top menu click on the Pivot Point tab. You should be able to see a beam is circular.	
14.	The current magnification is in the medium range.	46,000 x
15.	Make sure the circle is in the middle. Click on the Amplitude + Phase button on the top. Wait several seconds till it greys out.	This will try to bring the beam to a spot automatically.
16.	Click View and then click Offset Correction	This will try to bring the beam to one point with the precession on/off.
17.	Gradually go to higher magnification and at mag. 245,000x repeat the two steps above.	The beam may move by going to higher mag. Use trackball to bring the beam to the middle.
18.	Go to the diffraction mode on TEM control panel, then click on the Diff Focus tab in Topspin	
19.	Click View. Use magnification knob and change the Camera Length to 260 mm (or smaller) Change the cameral length of the software in the left menu, accordingly.	If the beam is off center, use Multifunction X and Y to bring the beam to the center.



20.	Click on the Series Area tab. Then click on Diffraction button. This is a preview scan tab that shows you where it is going to scan and how your sample looks like. The STEM Magnification on the bottom of the screen defines the preview scan magnification	
21.	By clicking on the scanning button, it starts showing you the preview. The estimate time is indicated on the left.	Changing the Frame Size define the number of scanning points in one line. More frame size, better resolusion, longer time.
22.		
23.	Click on the Focus tab. Press the View button to see the diffraction pattern. Normally no change is necessary at this point. However, advanced Focus tuning is explained.	Advanced Focus tuning: Use the blue arrow and put it on a feature with relatively high contrast. Click on Diffraction, then Beam Focus button (unprocessed profile). Click on Pivot Height and adjust the BDLX or BDLY Amplitude using the left alignment knobs on the Digistar hand panel to make the STEM profile with precession most closely match the unprecessed profile.
24.	You can skip the Setup Drift and go directly to Series tab.	Drift Correction is not necessary on this microscope. Sometimes with Drift Correction on, the acquition time may takes way longer.



25.	In the Series tab you can select the area for the final data acquisition (blue rectangle), the step size (resolution in nm) and number of precession per frame (diffraction pattern). 2 nm to 4 nm is the resolution of data acquisition for the Talos microscope.	
26.	Click Preview and put the blue circle in the diffraction pattern around the central beam (this will make the virtual bright field image)	The estimate time as shown on the left menu. 20 minutes to 30 minutes scanning time is normal.
27.	After the scan is done, in case your scan is satisfactory and you want to save your data, click on Archive .	
28.	On the desktop, open the application Topspin Data Viewer.	DigiStar Control
29.	Select Show Archived to find your scan. Double click on the image to open it.	OPSPIN Data Viewer Search Recomp Wandword Help Clear All Q Date 08/31/2018 to Export Archive Delete Import Show Archive All Export Archive Delete Import Show Archive All Export Archive Delete Import Show Archive All Export OP/06/2018 15:21 Experiment PED STEM/Series/Acquire Speciment PED STEM/Series/Acquire Speciment OP/06/2018 15:52 Date 09/06/2018 16:52 Date 08/31/2018 19:19-AlMn1(200xy,c) Experiment PO0500 Uset Date 09/06/2018 18:56 Date 08/31/2018 18:56 Date 09/06/2018 18:56-AlMn-1(326xy) Experiment PEO STEM/Series/Preview Speciment PEO STEM/Series/Acquire Speciment POS STEM/Series/Preview Speciment PEO STEM/Series/Preview Speciment PEO STEM/Series/Acquire Experiment POS STEM/Series/Preview Speciment PEO STEM/Series/Preview Speciment PEO STEM/Series/Acquire Speciment POS STEM/Series/Pre



		V TOPSPN bola Vener Santh Anomy Senth Anom
30.		🕼 08/31/2018 19.13-MMs12000y_2mmPix_1Prec_12min) 🕛 Powening * Antime Device
	icon (🍄) to correct the image	ASTABLOW Reference Virtual Sem Image Result Service
	gamma level to reveal some of	
	the faint spots in the diffraction	
	pattern.	And the second sec
		Vital Stem Image Result
	Then click ASTAR Export	
31.	Save your data in	Export Export South Biodefiles > + + South Biodefiles > P
	D:\Blockfiles\your personal folder	Organize • New folder Image: Constraint of the state of
		Convincads Allie 7/26/2018 3:18 PM File folder Amir 9/7/2018 3:37 PM File folder Amir 9/7/2018 3:37 PM File folder
	Saving this data may take several	Counters Converse
	minutes.	S. Pictures
		Computer Local Disk (C) Jord TA (D)
		AMBALI 1 (F) File name: 04.27.2018 15.06.54.bio
		nie name: 04,27,2008 i 32,06,5400 Save at type: [ASTAR (*blo)
		Hide Folders Save Cancel
32.		
	programs should be closed .	
	Data analysis and pattern solving	
	are described in separate SOP.	
33.		
55.	deactivate the NanoMEGAS and	
	FluCam in the NanoMegas tab.	
	Restart FluCam.	
34.	Close the TEM software as usual.	
25		
35.	BADGER LOGOUT: Don't	
	forget to disable the tool	
	in badger after you're	
	done.	

